

The Plasma Membrane Ca^{2+} -ATPase, a Molecular Target for Tau-induced Cytosolic Calcium Dysregulation

María Berrocal^{a,b} and Ana M. Mata^{a,b,*}

^a Departamento de Bioquímica y Biología Molecular y Genética, Facultad de Ciencias, Universidad de Extremadura, 06006 Badajoz, Spain

^b Instituto de Biomarcadores de Patologías Moleculares, Universidad de Extremadura, 06006 Badajoz, Spain

Abstract—Disruption of calcium (Ca^{2+}) homeostasis is emerging as a prevalent feature of aging and aging-associated neurodegenerative diseases, including Alzheimer's disease (AD), the most common type of tauopathy. This disease is characterized by the combined presence of extracellular neuritic plaques composed by amyloid β -peptides ($\text{A}\beta$) and neurofibrillary tangles of tau. The association of calcium dyshomeostasis with $\text{A}\beta$ has been extensively studied, however its link with tau has been less investigated. Thus, this review will concentrate on the functional link between tau and the plasma membrane Ca^{2+} pump (PMCA) and other membrane proteins involved in the regulation of intracellular calcium and/or its association with neurodegeneration.

This article is part of a Special Issue entitled: *Tauopathies*. © 2022 The Author(s). Published by Elsevier Ltd on behalf of IBRO. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Key words: PMCA, tau, Alzheimer's disease, calmodulin, methylene blue, sorcin.

INTRODUCTION

Transient increase in intracellular Ca^{2+} concentration is a major signaling event leading to the activation of many cellular processes (Berridge, 2013). But, in the non-activated state, the levels of cytosolic Ca^{2+} must be kept low (nM range) to preserve the cell function, and this is performed by the cell through several mechanisms. Among them, Ca^{2+} transport ATPases located in the plasma membrane (PMCA) and in the sarco/endoplasmic reticulum (SERCA) and secretory pathway (SPCA) play a major role in controlling the concentration of free Ca^{2+} in the cytosol. Disruption of neuronal Ca^{2+} homeostasis involves deregulation of these pumps and other Ca^{2+} -transporters and channels, and Ca^{2+} binding proteins.

Dysregulation of intracellular calcium (Ca^{2+}) is a common feature of many neurodegenerative pathologies and is one of the main sources of neuronal dysfunction in Alzheimer's disease (AD) (Magi et al., 2016). AD is characterized by the presence of aberrant aggregates of the amyloid- β peptide ($\text{A}\beta$) and phosphorylated tau. It is the most common tauopathy and more precisely it can be considered as a secondary tauopathy because the primary driver of the disease is the $\text{A}\beta$ peptide (Selkoe and Hardy, 2016; Chung et al., 2021). Several reviews highlight the involvement of PMCA and other proteins associ-

ated to Ca^{2+} signaling (such as the $\text{Na}^+/\text{Ca}^{2+}$ exchanger, plasma membrane Ca^{2+} channels, plasma membrane and endoplasmic reticulum receptors, Ca^{2+} buffers and Ca^{2+} sensors) in AD and other diseases associated to neuronal degeneration (Brini et al., 2014; Brini et al., 2017; Stafford et al., 2017; Hajjeva et al., 2018; Mata, 2018; Strehler and Thayer, 2018; O'Day, 2020; Boczek et al., 2021; Hwang et al., 2021).

Although tau is mostly a cytoplasmic protein bound to microtubules, it can also be localized at the cell membrane in pheochromocytoma (Brandt et al., 1995) and neuroblastoma cells (Arrasate et al., 2000), and in cerebral cortex affected by AD (Gray et al., 1987). Besides, Arrasate et al. (2000) reported that in neuroblastoma cells this localization is facilitated by dephosphorylation of tau in its proline-rich region, and suggested that this can be important in tauopathies, such as AD.

It has been widely reported that tau interacts with the cell membrane through its binding to lipids or/and membrane proteins, causing cellular dysfunctions and propagation of tauopathies. The N-terminal acidic projection domain and the C-terminal microtubule-binding domain of tau seem to play a key role in its interaction with the membrane and with membrane proteins (Brandt et al., 1995; Arrasate et al., 2000). Extensive reviews describe in full detail these interactions (Brandt et al., 2020; Brunello et al., 2020; Bok et al., 2021; Sallaberry et al., 2021). However, there are not many evidences about the interaction of tau with proteins involving alteration of their roles as modulators of Ca^{2+} homeostasis. This review provides an overview of the

*Corresponding author. Address: Departamento de Bioquímica y Biología Molecular y Genética, Facultad de Ciencias, Universidad de Extremadura, 06006 Badajoz, Spain.

E-mail address: anam@unex.es (A. M. Mata).

Abbreviations: AD, Alzheimer's disease; CaM, calmodulin; MB, methylene blue; PMCA, plasma membrane Ca^{2+} -ATPase.

relationship between tau and Ca^{2+} , focusing on the interaction of tau with several proteins. First, we summarize and discuss the main findings of our laboratory regarding the inhibitory effects of tau on the functioning of plasma membrane Ca^{2+} -ATPase (PMCA), which plays a major role in the fine tuning of intracellular Ca^{2+} , and the modulation of this inhibition, which may contribute to restoration of intracellular Ca^{2+} levels. In the second part we provide a brief review of tau binding with other major membrane-associated proteins, linked to tau pathologies.

FUNCTIONAL IMPAIRMENT OF PMCA BY TAU

A recent work by [Shrivastava et al. \(2019\)](#) pointed out the interaction of exogenous fibrillar tau with the neuron plasma membrane. In fact, they show that fibrillar tau clamps at the plasma membrane following lateral diffusion. By proteomic screening, using liquid chromatography and tandem mass spectrometry, these authors identified the interaction of exogenous tau with 29 membrane proteins. Among those interacting proteins they revealed the interaction of tau with plasma membrane Ca^{2+} -ATPase isoforms PMCA1 and PMCA4. Other authors ([Drummond et al., 2020](#)) also used affinity purification mass-spectrometry to identify a great number of membrane proteins that interact with tau. Seventy five proteins were associated to phosphorylated tau and 34 proteins were associated with total tau, including plasma membrane and intracellular Ca^{2+} -ATPases isoforms PMCA1 and SERCA2, respectively, and other ATPases such as the vacuolar ATPase, ATP synthase alpha/beta family proteins, V-type proton ATPase subunits D1, B2 and H and the Na^+ - K^+ -ATPase (subunits $\alpha 1$, $\alpha 2$ and $\beta 1$).

In line with Ca^{2+} dysregulation linked to neurodegeneration and specifically to AD, we have shown that among the three types of Ca^{2+} -ATPases, only PMCA is functionally affected by two main components of the histopathological marks of AD, the A β peptide ([Berrocal et al., 2009](#)) and tau ([Berrocal et al., 2015](#)). The peptide binds to a calmodulin (CaM) binding site which is present in the PMCA ([Berrocal et al., 2012](#)), but neither in SERCA nor in SPCA. Interestingly, the inhibitory effect of tau on PMCA was only seen when the pump was reconstituted with acidic phospholipids, while neutral lipids protected the protein from its inhibition by tau ([Berrocal et al., 2017](#)), suggesting that tau effects are highly dependent on the ionic nature of the phospholipids surrounding the protein.

Taking into consideration that both, the N- and the C-terminal regions of tau are involved in its interaction with the membrane and membrane proteins, and that the N terminus is negatively charged whereas the C terminal tail is mainly positively charged ([Rosenberg et al., 2008](#)), and also the charge of the substrates of PMCA (Ca^{2+} , H^+ , Mg^{2+} and ATP), we suggested that tau-PMCA binding is supported by electrostatic interactions, and that tau may use its N-terminus to interact through the plasma membrane with positively charged residues of PMCA. Activity assays performed at increasing concentrations of KCl showed a reversion of the inhibitory effect of tau on PMCA activity ([Berrocal et al., 2015](#)). Salts

may affect the interaction between water and protein side chains or backbone by masking charged residues which are important for PMCA binding to tau. Therefore, we can consider that ionic interactions play an important role in tau-PMCA binding. Furthermore, overlay assays led us to suggest that PMCA binding sites for tau should be located somewhere at its C-terminal cytosolic tail, and that it may involve the calmodulin binding site, or another region close to it. This was supported by further studies showing that tau impaired to some extent the binding of CaM to PMCA ([Berrocal et al., 2017](#)). In addition, we must consider membrane lipids as another factor that could contribute to the inhibitory effect of tau on PMCA.

The affinity of tau for lipids is dependent on its electrostatic interactions with phospholipids headgroup ([Künze et al., 2012](#)). In fact, tau binds preferentially to acidic phospholipids ([Yamauchi et al., 1997](#); [Jones et al., 2012](#); [Majewski et al., 2020](#)) and other charged lipid membranes ([Jones et al., 2012](#)), with dissociation constant values up to about 250 lower than for neutral lipid membranes ([Künze et al., 2012](#)). Therefore, the ionic nature of membrane lipids is relevant for the effects of tau on membrane proteins. It has been extensively reported that PMCA can be activated by acidic phospholipids such as PS, reaching the maximal activity ([Brodin et al., 1992](#); [Salvador and Mata, 1996](#)). Then, it is possible that the interaction of tau with acidic lipids could counteract the activating effect of these lipids on PMCA. Our studies corroborated this possibility as part of the mechanisms involved in the inhibition of PMCA by tau ([Berrocal et al., 2017](#)).

The works reviewed here have led us to propose that the failure of PMCA function by tau points out the involvement of PMCA in pathologies such as AD and related tauopathies.

COUNTERACTING THE EFFECTS OF TAU ON PMCA BY CALMODULIN, SORCIN AND METHYLENE BLUE

We focused our interest to identify compounds that could block or prevent the negative effects of tau on PMCA functioning. Up to now we have found beneficial effects with two proteins, calmodulin (CaM) and sorcin, and with the methylene blue (MB) dye. [Fig. 1](#) summarize the strategies followed by those compounds to counteract the inhibitory effect of tau on PMCA activity.

Calmodulin and PMCA

CaM is the main endogenous activator of PMCA. In the presence of Ca^{2+} , CaM binds to the CaM binding site of the pump, which also serves as an autoinhibitor, and releases the protein from the autoinhibition ([Carafoli, 1997](#)). As a result, the PMCA became fully activated and then is able to pump the excess of cytosolic Ca^{2+} out of the cell, thus maintaining the optimum low intracellular Ca^{2+} levels. We have already reported that CaM decreases and even prevents the inhibition of PMCA activity by tau ([Berrocal et al., 2017](#)). Although the inhibitory effect of tau does not interfere with the activating effect of CaM on PMCA, tau reduced slightly the binding

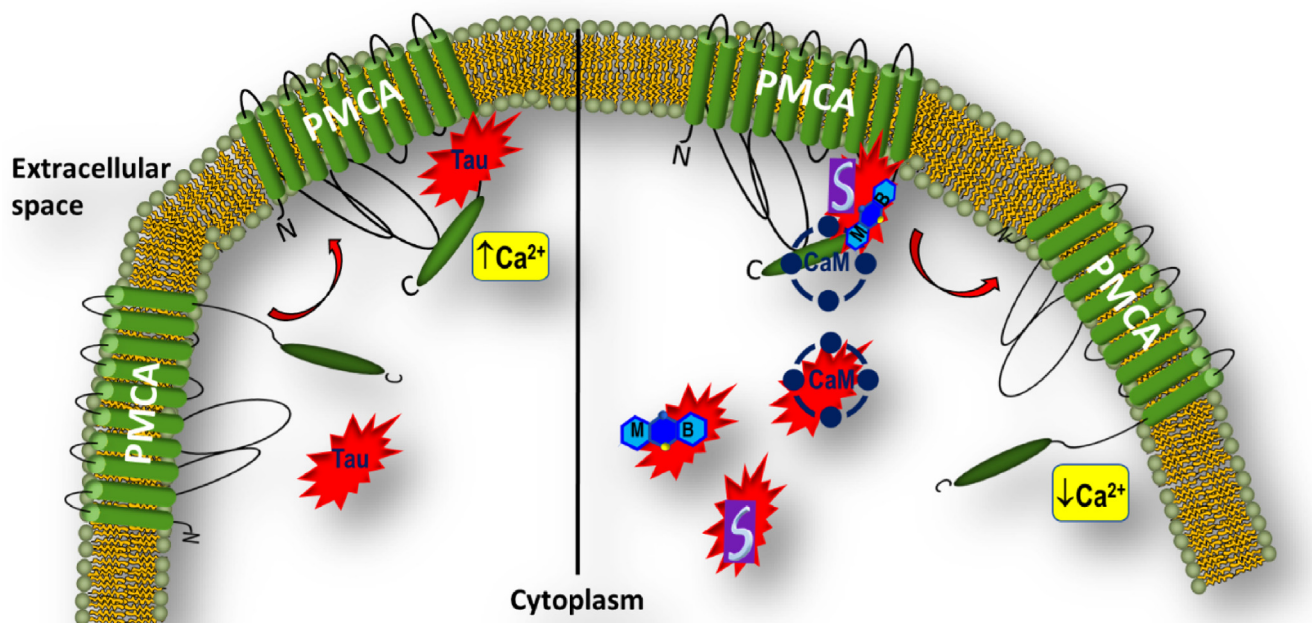


Fig. 1. Summary of the effects of tau and blocking agents on PMCA activity. The left side of the figure shows the inhibition of PMCA by tau binding, leading to cytosolic Ca^{2+} increase. The right side of the figure shows that calmodulin (CaM), sorcin (S) and methylene blue (MB) interact with both PMCA and tau, reversing or blocking its toxic effect, and then restoring PMCA activity.

of CaM to PMCA, showing a 30% increase in the Kd. We also found competition between inhibitory and protective effects produced by tau and CaM, respectively, on Ca^{2+} -ATPase activity. But an equimolar tau/CaM ratio was enough to prevent the inhibitory action of tau (Berrocal et al., 2015; Berrocal et al., 2017). Besides, overlay interaction assays showed that tau not only bind to PMCA but also to CaM. Therefore, an increase in CaM concentration results in a decrease of free tau to inhibit PMCA (Berrocal et al., 2017).

Sorcin and PMCA

The soluble resistance-related Ca^{2+} -binding protein (sorcin) has also been described by our group as a blocking agent of the inhibitor effect of tau on PMCA. Data bank report that sorcin is highly expressed in brain (Andreev et al., 2012; Hondius et al., 2018; Sathe et al., 2021). Further studies have shown a molecular interaction of sorcin with tau in cell cultures overexpressing both proteins (Kim et al., 2016), with presenilin-2 (which is involved in the onset of familial AD) in cell cultures and human brain tissues (Pack-Chung et al., 2000), and also with synuclein and synphilin-1 (involved in Parkinson's disease and other Lewy body diseases) using phage display (Woods et al., 2007) and gene co-expression analysis (George et al., 2019). Besides, this protein activates the cardiac SERCA pump (Matsumoto et al., 2005). Those finding lead us to investigate the association of sorcin with PMCA. By using kinetic approaches we found that sorcin was able to activate all SERCA isoforms, and also the PMCA pump in purified preparations of pig brain, in human brain membranes and in COS cell mem-

branes overexpressing each of the four main PMCA isoforms (Berrocal et al., 2021). Besides, it could block the inhibitory effects of tau in all preparations. Sorcin was also able to prevent the toxicity of tau in human SH-SY5Y cells, as showed by cell viability, ROS production and apoptosis assays. Additional binding assays were carried out to demonstrate that functional effects were associated to molecular interactions of sorcin, not only with the PMCA but also with tau. In addition, sorcin was able to activate the truncated variant hPMCA4b-R1052* lacking the whole C-terminal tail, that resembles the structure of SERCA, suggesting that sorcin may bind to a similar region or pocket in both calcium pumps. Thus, the interaction of sorcin with tau could be a significant mechanism to avoid the partial or total loss of PMCA function by tau. Similar effects were also reported for sorcin and A β (Berrocal et al., 2021).

Methylene blue and PMCA

The phenothiazine MB is a dye which has been widely used in several fields including chemistry, medicine or as a photosensitizing agent to treat bacterial and fungal infections. MB attracted our attention because a growing number of evidence have shown neuroprotective roles of MB against brain damage in mice and rats (Mori et al., 2014; Huang et al., 2018; Cheng et al., 2021), and also in patients with mild AD (Wilcock et al., 2018). In fact, a stable reduced form of the MB (leucomethylene blue), acts as a selective inhibitor of tau aggregation, both in vitro and in transgenic mouse models, and it has been used in clinical trials in patients with mild to moderate AD (Wilcock et al., 2018; Schelter et al., 2019). As pointed out

above, the general idea is that the beneficial effect of tau is due to inhibition of tau aggregation, preventing the toxic effects of tau. However, it has been shown that the monomeric form of tau is sufficient to initiate the spread of tau pathology (Michel et al., 2014), and dimers and trimers could also induce toxicity (Cowan and Mudher, 2013). We then, performed functional studies with tau and PMCA in the presence of MB to see if this phenothiazine could exert a protective effect against tau toxicity through its interaction with PMCA. By using purified PMCA and crude membranes from human tissues affected by AD and control subjects and from neuroblastoma cells, MB could block and even reverse the inhibitory effect of tau on PMCA activity (Berrocal et al., 2019). Besides, MB was able to protect neuroblastoma cell cultures against DNA damage due to loss of cell viability and increase of ROS production, due from exogenous tau. Fluorescence experiments allow us to propose that MB interacts and binds to PMCA at micromolar concentrations, and that this interaction was modulated by tau. In fact, the quenching of intrinsic fluorescence of PMCA by MB was largely increased in the presence of tau, suggesting that tau significantly increases the affinity of MB for PMCA, in such way that just micromolar MB concentrations will be needed to bind the brain PMCA in cells. We then proposed that this drug may have beneficial effects in AD by preventing and blocking the toxic effects of tau on PMCA function associated to the maintenance of cellular Ca^{2+} homeostasis (Berrocal et al., 2019).

INTERACTION OF TAU WITH OTHER MEMBRANE PROTEINS RELATED TO Ca^{2+} DYSREGULATION AND/OR TAUOPATHIES

The relationship between tau and Ca^{2+} dysregulation has been revealed in cell cultures and in transgenic tauopathy mice models. It has been reported that extracellular tau induces an increase of intracellular Ca^{2+} concentrations in SH-SY5Y neuroblastoma cells and in COS-7 non-neuronal cells, leading to toxic effects (Gomez-Ramos et al., 2006; Gómez-Ramos et al., 2008). This increase has been attributed to its selective interaction with **M1 and M3 muscarinic receptors**, that promote the release of Ca^{2+} from intracellular compartments. By performing fluorescence assays in Fura-2 loaded cells transfected with M1 and M3 cDNAs in the presence of tau, acetylcholine and atropine, as agonist and antagonist of muscarinic receptors, respectively, they found that the increase of cytosolic Ca^{2+} induced by acetylcholine was suppressed by atropine, indicating that muscarinic receptors were the main cholinergic receptors in that cell line. Besides, extracellular tau induced the same effect as acetylcholine, suggesting that tau mediates the increase in Ca^{2+} concentration through its interaction with muscarinic receptors. The Ca^{2+} concentration increase was not seen after addition of atropine (Gómez-Ramos et al., 2008). Both muscarinic receptors are highly expressed in the hippocampus and entorhinal cortex of mice, two areas involved in tau pathology at the early stages of the AD. Besides, other proteins related to neurodegeneration also produce dysregulation of Ca^{2+}

homeostasis in neuronal cell cultures (Demuro et al., 2005; Danzer et al., 2007).

Other studies (Smith et al., 1995; Giaccone et al., 1996; Islam and Levy, 1997), have highlighted the interaction of tau with the **amyloid precursor protein (APP)**, which plays a key role in AD pathogenesis, because it is the precursor of toxic A β peptides, and its mutations are linked to familial AD. However, the amino acid residues of APP involved in its binding with tau are under controversy (Maron et al., 2020; Britton et al., 2021). It has also been shown (Piacentini et al., 2017) that APP may act as a receptor of extracellular tau in astrocytes, inducing its internalization and subsequent impairment of intracellular Ca^{2+} signaling that is essential for gliotransmitter release.

The **apoE4** is the strongest genetic risk factor for late-onset AD. It has been shown that APOE4 produces a considerable reduction of protein synthesis in neurons and a continuous increase in Ca^{2+} levels by activating both NMDARs and L-type voltage-gated calcium channels. Thus, disruption of Ca^{2+} homeostasis by APOE4 leads to a defective protein synthesis. These observations could explain the APOE4-mediated predisposition to AD and involve Ca^{2+} dyshomeostasis in the molecular events associated to ApoE4 and AD (Ramakrishna et al., 2021). Other studies have reported that this isoform aggravates A β -mediated neurodegeneration by its interaction with the neuronal low-density lipoprotein receptor-related protein 1 (**LRP1**) (Tachibana et al., 2019). Recently, LRP1, has been identified as an endocytic receptor for extracellular tau (Rauch et al., 2020). Other authors have reported that LRP1 binds with lower affinity to phosphorylated tau and then is less efficiently internalized by LRP1 (Cooper et al., 2021). Besides, the LRP1-mediated uptake of tau is inhibited by apoE4. It has also been reported that ApoE4 affects tau pathogenesis, tau-mediated neurodegeneration and neuroinflammation independently of A β pathology (Shi et al., 2017). Thus, the authors propose ApoE4 as a potential therapeutic approach in reducing tau-mediated neurodegeneration and suggest a re-evaluation of the role of ApoE in AD and other tauopathies.

The interaction of tau with these proteins may lead to several effects, depending on the proteins and tau species, and on the cell type. It is widely known that aggregates of hyperphosphorylated tau in the AD brain are a characteristic hallmark of AD. However, it has also been shown that soluble monomers and dimers of extracellular tau are highly toxic to hippocampal granule neurons (Bolós et al., 2017; Bengoa-Vergniory et al., 2021). Wu et al. (2021), have also reported that Ca^{2+} dyshomeostasis correlates better with soluble than insoluble pathological tau in mouse models of tauopathy. Besides, Perea et al. (2018) have shown that dephosphorylated rather than hyperphosphorylated tau generates a pro-inflammatory response in microglia through the activation of p38 mitogen-activated protein kinase (**MAPK**) pathway. And more recently, the same group have reported that this toxic effect of tau can be reversed by pharmacological inhibition of P38 (Perea et al., 2022). As a result, they suggest that P38 inhibition can be a potential therapeutic strategy for tauopathies.

In summary, Ca²⁺ dysregulation associated to tauopathies such as AD, could be mediated by the interaction of tau with membrane proteins such as the PMCA and other key proteins involved in neurodegeneration.

ACKNOWLEDGEMENTS

We thank past and present members of the laboratory and colleagues who have contributed to some of the studies described in this review. These studies have been supported by Projects BFU2014-53641-P, BFU2017-85723-P and PID2020-115512 GB-I00, funded by MCIN/AEI/10.13039/501100011033 and by “ESF Investing in your future”.

REFERENCES

- Andreev VP, Petyuk VA, Brewer HM, Karpievitch YV, Xie F, Clarke J, Camp D, Smith RD, Lieberman AP, Albin RL, Nawaz Z, El Hokayem J, Myers AJ (2012) Label-free quantitative LC-MS proteomics of Alzheimer's disease and normally aged human brains. *J Proteome Res* 11(6):3053–3067.
- Arrasate M, Pérez M, Avila J (2000) Tau dephosphorylation at tau-1 site correlates with its association to cell membrane. *Neurochem Res* 25:43–50. <https://doi.org/10.1023/a:1007583214722>.
- Bengoa-Vergniory N, Velentza-Almpani E, Silva AM, Scott C, Vargas-Caballero M, Sastre M, Wade-Martins R, Alegre-Abarrategui J (2021) Tau-proximity ligation assay reveals extensive previously undetected pathology prior to neurofibrillary tangles in preclinical Alzheimer's disease. *acta neuropathol commun* 9(1).
- Berridge MJ (2013) Calcium regulation of neural rhythms, memory and Alzheimer's disease. *J Physiol* 592:281–293. <https://doi.org/10.1113/jphysiol.2013.257527>.
- Berrocal M, Marcos D, Sepúlveda MR, Pérez M, Avila J, Mata AM (2009) Altered Ca²⁺ dependence of synaptosomal plasma membrane Ca²⁺-ATPase in human brain affected by Alzheimer's disease. *FASEB J* 23:1826–1834. <https://doi.org/10.1096/fj.08-121459>.
- Berrocal M, Sepúlveda MR, Vazquez-Hernandez M, Mata AM (2012) Calmodulin antagonizes amyloid- β peptide-mediated inhibition of brain plasma membrane Ca(2+)-ATPase. *Biochim Biophys Acta* 1822:961–969. <https://doi.org/10.1016/j.bbadis.2012.02.013>.
- Berrocal M, Corbacho I, Vázquez-Hernández M, Ávila J, Sepúlveda MR, Mata AM (2015) Inhibition of PMCA activity by tau as a function of aging and Alzheimer's neuropathology. *Biochim Biophys Acta* 1852:1465–1476. <https://doi.org/10.1016/j.bbadis.2015.04.007>.
- Berrocal M, Corbacho I, Sepúlveda MR, Gutierrez-Merino C, Mata AM (2017) Phospholipids and calmodulin modulate the inhibition of PMCA activity by tau. *Biochim Biophys Acta - Molecular Cell Research* 1864:1028–1035. <https://doi.org/10.1016/j.bbamcr.2016.10.023>.
- Berrocal M, Caballero-Bermejo M, Gutierrez-Merino C, Mata AM (2019) Methylene Blue Blocks and Reverses the Inhibitory Effect of Tau on PMCA Function. *Int J Mol Sci* 20:3521. <https://doi.org/10.3390/ijms20143521>.
- Berrocal M, Saez L, Mata AM (2021) Sorcin Activates the Brain PMCA and Blocks the Inhibitory Effects of Molecular Markers of Alzheimer's Disease on the Pump Activity. *Int J Mol Sci* 22:6055. <https://doi.org/10.3390/ijms22116055>.
- Boczek T, Sobolczyk M, Mackiewicz J, Lisek M, Ferenc B, Guo F, Zylinska L (2021) Crosstalk among Calcium ATPases: PMCA, SERCA and SPCA in Mental Diseases. *Int J Mol Sci* 22:2785. <https://doi.org/10.3390/ijms22062785>.
- Bok E, Leem E, Lee BR, Lee JM, Yoo CJ, Lee EM, Kim J (2021) Role of the Lipid Membrane and Membrane Proteins in Tau Pathology. *Front Cell Dev Biol* 9. <https://doi.org/10.3389/fcell.2021.653815>.
- Bolós M, Pallas-Bazarra N, Terreros-Roncal J, Perea JR, Jurado-Arjona J, Ávila J, Llorens-Martín M (2017) Soluble tau has devastating effects on the structural plasticity of hippocampal granule neurons. *Transl Psychiatry* 7(12). <https://doi.org/10.1038/s41398-017-0013-6>.
- Brandt R, Léger J, Lee G (1995) Interaction of tau with the neural plasma membrane mediated by tau's amino-terminal projection domain. *J Cell Biol* 131:1327–1340. <https://doi.org/10.1083/jcb.131.5.1327>.
- Brandt R, Trushina NI, Bakota L (2020) Much More Than a Cytoskeletal Protein: Physiological and Pathological Functions of the Non-microtubule Binding Region of Tau. *Front Neurol* 11. <https://doi.org/10.3389/fneur.2020.590059>.
- Brini M, Cali T, Ottolini D, Carafoli E (2014) Neuronal calcium signaling: function and dysfunction. *Cell Mol Life Sci* 71:2787–2814. <https://doi.org/10.1007/s00018-013-1550-7>.
- Brini M, Carafoli E, Cali T (2017) The plasma membrane calcium pumps: focus on the role in (neuro)pathology. *Biochem Biophys Res Commun* 483:1116–1124. <https://doi.org/10.1016/j.bbrc.2016.07.117>.
- Britton RJ, Hutchison JM, Sanders CR (2021) The transmembrane amyloid precursor C99 protein exhibits non-specific interaction with tau. *Biochem Biophys Res Commun* 576:48–52. <https://doi.org/10.1016/j.bbrc.2021.08.075>.
- Brodin P, Falchetto R, Vorherr T, Carafoli E (1992) Identification of two domains which mediate the binding of activating phospholipids to the plasma-membrane Ca²⁺ pump. *Eur J Biochem* 204:939–946. <https://doi.org/10.1111/j.1432-1033.1992.tb16715.x>.
- Brunello CA, Merezko M, Uronen RL, Huttunen HJ (2020) Mechanisms of secretion and spreading of pathological tau protein. *Cell Mol Life Sci* 77:1721–1744. <https://doi.org/10.1007/s00018-019-03349-1>.
- Carafoli E (1997) Plasma membrane calcium pump: structure, function and relationships. *Basic Res Cardiol* 92:59–61. <https://doi.org/10.1007/BF00794069>.
- Cheng Q, Chen X, Ma J, Jiang X, Chen J, Zhang M, Wu Y, Zhang W, Chen C, Ostrowski R (2021) Effect of Methylene Blue on White Matter Injury after Ischemic Stroke. *Oxidative Medicine and Cellular Longevity* 2021:1–10.
- Chung DeC, Roemer S, Petrucelli L, Dickson DW (2021) Cellular and pathological heterogeneity of primary tauopathies. *Mol Neurodegeneration* 16:57. <https://doi.org/10.1186/s13024-021-00476-x>.
- Cooper JM, Lathuiliere A, Migliorini M, Arai AL, Wani MM, Dujardin S, Muratoglu SC, Hyman BT, Strickland DK (2021) Regulation of tau internalization, degradation, and seeding by LRP1 reveals multiple pathways for tau catabolism. *J Biol Chem* 296. <https://doi.org/10.1016/j.jbc.2021.100715>.
- Cowan CM, Mudher A (2013) Are tau aggregates toxic or protective in tauopathies? *Front Neurol* 4:114. <https://doi.org/10.3389/fneur.2013.00114>.
- Danzer KM, Haasen D, Karow AR, Moussaud S, Habeck M, Giese A, Kretschmar H, Hengerer B, Kostka M (2007) Different species of alpha-synuclein oligomers induce calcium influx and seeding. *J Neurosci* 27:9220–9232. <https://doi.org/10.1523/JNEUROSCI.2617-07.2007>.
- Demuro A, Mina E, Kaye R, Milton SC, Parker I, Glabe CG (2005) Calcium dysregulation and membrane disruption as a ubiquitous neurotoxic mechanism of soluble amyloid oligomers. *J Biol Chem* 280:17294–17300. <https://doi.org/10.1074/jbc.M500997200>.
- Drummond E, Pires G, MacMurray C, Askenazi M, Nayak S, Bourdon M, Safar J, Ueberheide B, Wisniewski T (2020) Phosphorylated tau interactome in the human Alzheimer's disease brain. *Brain* 143:2803–2817. <https://doi.org/10.1093/brain/awaa223>.
- George G, Singh S, Lokappa SB, Varkey J (2019) Gene co-expression network analysis for identifying genetic markers in Parkinson's disease - a three-way comparative approach.

- Genomics 111:819–830. <https://doi.org/10.1016/j.ygeno.2018.05.005>.
- Giaccone G, Pedrotti B, Migheli A, Verga L, Perez J, Racagni G, Smith MA, Perry G, et al. (1996) beta PP and Tau interaction. A possible link between amyloid and neurofibrillary tangles in Alzheimer's disease. *Am J Pathol* 48:79–87. PMID: 8546229.
- Gomez-Ramos A, Diaz-Hernandez M, Cuadros R, Hernandez F, Avila J (2006) Extracellular tau is toxic to neuronal cells. *Febs Lett* 580:4842–4850. <https://doi.org/10.1016/j.febslet.2006.07.078>.
- Gómez-Ramos A, Díaz-Hernández M, Rubio A, Miras-Portugal MT, Avila J (2008) Extracellular tau promotes intracellular calcium increase through M1 and M3 muscarinic receptors in neuronal cells. *Mol Cell Neurosci* 37:673–681. <https://doi.org/10.1016/j.mcn.2007.12.010>.
- Gray EG, Paula-Barbosa M, Roher (1987) Alzheimer's disease: paired helical filaments and cytomembranes. *Neuropathol Appl Neurobiol* 13:91–110. <https://doi.org/10.1111/j.1365-2990.1987.tb00174.x>.
- Hajjeva P, Baeken MW, Moosmann B (2018) The role of Plasma Membrane Calcium ATPases (PMCA) in neurodegenerative disorders. *Neurosci Lett* 663:29–38. <https://doi.org/10.1016/j.neulet.2017.09.033>.
- Hondius DC, Eigenhuis KN, Morrema THJ, van der Schors RC, van Nierop P, Bugiani M, Li KW, Hoozemans JJM, Smit AB, Rozemuller AJM (2018) Proteomics analysis identifies new markers associated with capillary cerebral amyloid angiopathy in Alzheimer's disease. *Acta Neuropathol Commun* 6(1). <https://doi.org/10.1186/s40478-018-0540-2>.
- Huang L, Lu J, Cerqueira B, Liu Y, Jiang Z, Duong TQ (2018) Chronic oral methylene blue treatment in a rat model of focal cerebral ischemia/reperfusion. *Brain Res* 1678:322–329. <https://doi.org/10.1016/j.brainres.2017.10.033>.
- Hwang SM, Lee JY, Park CK, Kim YH (2021) The Role of TRP Channels and PMCA in Brain Disorders: Intracellular Calcium and pH Homeostasis. *Front Cell Dev Biol* 9. <https://doi.org/10.3389/fcell.2021.584388> 584388.
- Islam K, Levy E (1997) Carboxyl-terminal fragments of beta-amyloid precursor protein bind to microtubules and the associated protein tau. *Am J Pathol* 151:265–71. PMID: 9212751.
- Jones EM, Dubey M, Camp PJ, Vernon BC, Biernat J, Mandelkow E, Majewski J, Chi EY (2012) Interaction of Tau Protein with Model Lipid Membranes Induces Tau Structural Compaction and Membrane Disruption. *Biochemistry* 51(12):2539–2550.
- Kim SI, Lee HJ, Kim SS, Kwon YS, Chun W (2016) Sequestration of sorcin by aberrant forms of tau results in the defective calcium homeostasis. *Korean J Physiol Pharmacol* 20:387–397. <https://doi.org/10.4196/kjpp.2016.20.4.387>.
- Künze G, Barré P, Scheidt HA, Thomas L, Eliezer D, Huster D (2012) Binding of the three-repeat domain of tau to phospholipid membranes induces an aggregated-like state of the protein. *Biochim Biophys Acta* 1818:2302–2313. <https://doi.org/10.1016/j.bbamem.2012.03.019>.
- Magi S, Castaldo P, Macri ML, Maiolino M, Matteucci A, Bastioli G, Gratteri S, Amoroso S, Lariccia V (2016) Intracellular Calcium Dysregulation: Implications for Alzheimer's Disease. *Biomed Res Int* 2016:6701324. <https://doi.org/10.1155/2016/6701324>.
- Majewski J, Jones EM, Vander Zanden CM, Biernat J, Mandelkow E, Chi EY (2020) Lipid membrane templated misfolding and self-assembly of intrinsically disordered tau protein. *Sci Rep* 10:13324. <https://doi.org/10.1038/s41598-020-70208-6>. PMID: 32770092; PMCID: PMC7414892.
- Maron R, Armony G, Tsoory M, Wilchek M, Frenkel D, Arnon R (2020) Peptide Interference with APP and Tau Association: Relevance to Alzheimer's Disease Amelioration. *Int J Mol Sci* 21:3270. <https://doi.org/10.3390/ijms21093270>.
- Mata AM (2018) Functional interplay between plasma membrane Ca²⁺-ATPase, amyloid β -peptide and tau. *Neurosci Lett* 663:55–59. <https://doi.org/10.1016/j.neulet.2017.08.004>.
- Matsumoto T, Hisamatsu Y, Ohkusa T, Inoue N, Sato T, Suzuki S, Ikeda Y, Matsuzaki M (2005) Sorcin interacts with sarcoplasmic reticulum Ca²⁺-ATPase and modulates excitation-contraction coupling in the heart. *Basic Res Cardiol* 100:250–262. <https://doi.org/10.1007/s00395-005-0518-7>.
- Michel CH, Kumar S, Pinotsi D, Tunnacliffe A, St George-Hyslop P, Mandelkow E, Mandelkow EM, Kaminski CF, Kaminski Schierle GS (2014) Extracellular monomeric tau protein is sufficient to initiate the spread of tau protein pathology. *J Biol Chem* 289:956–967. <https://doi.org/10.1074/jbc.M113.515445>.
- Mori T, Koyama N, Segawa T, Maeda M, Maruyama N, Kinoshita N, Hou H, Tan J, Town T (2014) Methylene blue modulates β -secretase, reverses cerebral amyloidosis, and improves cognition in transgenic mice. *J Biol Chem* 289:30303–30317. <https://doi.org/10.1074/jbc.M114.568212>.
- O'Day DH (2020) Calmodulin Binding Proteins and Alzheimer's Disease: Biomarkers, Regulatory Enzymes and Receptors That Are Regulated by Calmodulin. *Int J Mol Sci* 21:7344. <https://doi.org/10.3390/ijms21197344>.
- Pack-Chung E, Meyers MB, Pettingell WP, Moir RD, Brownawell AM, Cheng I, Tanzi RE, Kim TW (2000) Presenilin 2 interacts with sorcin, a modulator of the ryanodine receptor. *J Biol Chem* 275:14440–14445. <https://doi.org/10.1074/jbc.m909882199>.
- Perea JR, Ávila J, Bolós M (2018) Dephosphorylated rather than hyperphosphorylated Tau triggers a pro-inflammatory profile in microglia through the p38 MAPK pathway. *Exp Neurol* 310:14–21. <https://doi.org/10.1016/j.expneurol.2018.08.007>.
- Perea JR, Bolós M, Cuadros R, García E, García-Escudero V, Hernández F, McManus RM, Heneka MT, Avila J (2022) p38 Inhibition decreases tau toxicity in microglia and improves their phagocytic function. *Mol Neurobiol* 59(3):1632–1648.
- Piacentini R, Li Puma DD, Mainardi M, Lazzarino G, Tavazzi B, Arancio O, Grassi C (2017) Reduced gliotransmitter release from astrocytes mediates tau-induced synaptic dysfunction in cultured hippocampal neurons. *Glia* 65:1302–1316. <https://doi.org/10.1002/glia.23163>.
- Ramakrishna S, Jhaveri V, Konings SC, Nawalpur B, Chakraborty S, Holst B, Schmid B, Gouras GK, Freude KK, Muddashetty RS (2021) APOE4 affects basal and NMDAR-mediated protein synthesis in neurons by perturbing calcium homeostasis. *J Neurosci* 41(42):8686–8709.
- Rauch JN, Luna G, Guzman E, Audouard M, Challis C, Sibih YE, Leshuk C, Hernandez I, Wegmann S, Hyman BT, Gradinaru V, Kampmann M, Kosik KS (2020) LRP1 is a master regulator of tau uptake and spread. *Nature* 580(7803):381–385.
- Rosenberg KJ, Ross JL, Feinstein HE, Feinstein SC, Israelachvili J (2008) Complementary dimerization of microtubule-associated tau protein: Implications for microtubule bundling and tau-mediated pathogenesis. *Proc Natl Acad Sci U S A* 105:7445–7450. <https://doi.org/10.1073/pnas.0802036105>.
- Sallaberry CA, Voss BJ, Majewski J, Biernat J, Mandelkow E, Chi EY, Vander Zanden CM (2021) Tau and membranes: interactions that promote folding and condensation. *Front Cell Dev Biol* 9. <https://doi.org/10.3389/fcell.2021.725241> 725241.
- Salvador JM, Mata AM (1996) Purification of the synaptosomal plasma membrane (Ca²⁺ + Mg²⁺)-ATPase from pig brain. *Biochem J* 315:183–187. <https://doi.org/10.1042/bj3150183>.
- Sathe G, Albert M, Darrow J, Saito A, Troncoso J, Pandey A, Moghekar A (2021) Quantitative proteomic analysis of the frontal cortex in Alzheimer's disease. *J Neurochem* 156:988–1002. <https://doi.org/10.1111/jnc.15116>.
- Schelter BO, Shiells H, Baddeley TC, Rubino CM, Ganesan H, Hammel J, Vuksanovic V, Staff RT, Murray AD, Bracoud L, Riedel G, Gauthier S, Jia J, Benthham P, Kook K, Storey JMD, Harrington CR, Wischik CM (2019) Concentration-dependent activity of hydromethylthionine on cognitive decline and brain atrophy in mild to moderate Alzheimer's Disease. *J Alzheimers Dis* 72(3):931–946.
- Selkoe DJ, Hardy J (2016) The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med* 8:595–608. <https://doi.org/10.15252/emmm.201606210>.

- Shi Y, Yamada K, Liddelov SA, Smith ST, Zhao L, Luo W, Tsai RM, Spina S, Grinberg LT, Rojas JC, Gallardo G, Wang K, Roh J, Robinson G, Finn MB, Jiang H, Sullivan PM, Baufeld C, Wood MW, Sutphen C, McCue L, Xiong C, Del-Aguila JL, Morris JC, Cruchaga C, Fagan AM, Miller BL, Boxer AL, Seeley WW, Butovsky O, Barres BA, Paul SM, Holtzman DM (2017) ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. *Nature* 549(7673):523–527.
- Shrivastava AN, Redeker V, Pieri L, Bousset L, Renner M, Madióna K, hes-Hamon C, Coens A, et al. (2019) Clustering of Tau fibrils impairs the synaptic composition of α 3-Na⁺/K⁺-ATPase and AMPA receptors. *EMBO J* 38:e99871.
- Smith MA, Siedlak SL, Richey PL, Mulvihill P, Ghiso J, Frangione B, Tagliavini F, Giaccone G, et al. (1995) (1995) Tau protein directly interacts with the amyloid beta-protein precursor: implications for Alzheimer's disease. *Nat Med* 1:365–369. <https://doi.org/10.1038/nm0495-365>.
- Stafford N, Wilson C, Oceandy D, Neyses L, Cartwright EJ (2017) The Plasma Membrane Calcium ATPases and Their Role as Major New Players in Human Disease. *Physiol Rev* 97:1089–1125. <https://doi.org/10.1152/physrev.00028.2016>.
- Strehler EE, Thayer SA (2018) Evidence for a role of plasma membrane calcium pumps in neurodegenerative disease: Recent developments. *Neurosci Lett* 663:39–47. <https://doi.org/10.1016/j.neulet.2017.08.035>.
- Tachibana M, Holm M-L, Liu C-C, Shinohara M, Aikawa T, Oue H, Yamazaki Yu, Martens YA, Murray ME, Sullivan PM, Weyer K, Glerup S, Dickson DW, Bu G, Kanekiyo T (2019) APOE4-mediated amyloid- β pathology depends on its neuronal receptor LRP1. *J Clin Invest* 129(3):1272–1277.
- Wilcock GK, Gauthier S, Frisoni GB, Jia J, Hardlund JH, Moebius HJ, Bentham P, Kook KA, Schelter BO, Wischik DJ, Davis CS, Staff RT, Vuksanovic V, Ahearn T, Bracoud L, Shamsi K, Marek K, Seibyl J, Riedel G, Storey JMD, Harrington CR, Wischik CM (2018) Potential of Low Dose Leuco-Methylthionium Bis (Hydromethanesulphonate) (LMTM) Monotherapy for Treatment of Mild Alzheimer's Disease: Cohort Analysis as Modified Primary Outcome in a Phase III Clinical Trial. *J Alzheimers Dis* 61(1):435–457.
- Woods WS, Boettcher JM, Zhou DH, Kloepper KD, Hartman KL, Lador DT, Qi Z, Rienstra CM, George JM (2007) Conformation-specific binding of alpha-synuclein to novel protein partners detected by phage display and NMR spectroscopy. *J Biol Chem* 282:34555–34567. <https://doi.org/10.1074/jbc.M705283200>.
- Wu Q, Bai Y, Li W, Congdon EE, Liu W, Lin Y, Ji C, Gan WB, Sigurdsson EM (2021) Increased neuronal activity in motor cortex reveals prominent calcium dyshomeostasis in tauopathy mice. *Neurobiol Dis* 147. <https://doi.org/10.1016/j.nbd.2020.105165>.
- Yamauchi E, Titani K, Taniguchi H (1997) Specific binding of acidic phospholipids to microtubule-associated protein MAP1B regulates its interaction with tubulin. *J Biol Chem* 272:22948–22953. <https://doi.org/10.1074/jbc.272.36.22948>.

(Received 16 February 2022, Accepted 18 April 2022)
(Available online xxxx)