

1 **Repeated jumps from Northwest Africa to the European continent: the case of**  
2 **peripheral populations of an annual plant**

3 **Running title:** Peripheral populations of *Scrophularia arguta*

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15

1 **ABSTRACT**

2 Peripheral populations (i.e. those occurring on the edge of a species' distribution range)  
3 can have different origins and genetic characteristics, and they may be critical for  
4 conservation of genetic diversity. We investigated European peripheral populations of  
5 *Scrophularia arguta*, a widespread, annual plant distributed from Arabia to Northwest  
6 Africa and Macaronesia. Only two small disjunct population groups of this species occur  
7 in Europe, specifically in West-Central and Southeast Iberia. To disclose the origin of  
8 these populations and determine their importance for conservation of *S. arguta* genetic  
9 diversity, we analysed DNA sequences from two nuclear and two plastid regions and  
10 AFLP markers in populations sampled mainly across the western distribution range of the  
11 species, and modelled the species distribution under present and late Quaternary  
12 conditions. The analyses revealed the presence of three distinct lineages of *S. arguta* in  
13 Europe, as a result of multiple colonisation waves at different times in the Quaternary.  
14 Two of these lineages, occurring in Southeast Iberia, are the result of more or less recent  
15 dispersal from Northwest Africa. In contrast, West-Central Iberian populations are  
16 strongly differentiated from the remaining range of *S. arguta*, and can be considered as  
17 peripheral relict populations. Our study is the first to demonstrate the occurrence of at  
18 least three colonisations of the European continent from Africa by a native plant species.  
19 The diverse histories and genetic makeup of resulting populations confirm the importance  
20 of peripheral populations, and particularly of ancient relict populations, for conservation  
21 of global genetic diversity in widespread species.

22 **Key words:** Disjunct populations, genetic diversity, long-distance dispersal, migration  
23 routes, phylogeography, *Scrophularia*.

# 1. INTRODUCTION

Peripheral populations are defined as those populations occurring on the edge of a species' distribution range. Usually, they are smaller, more isolated and highly fragmented, have low effective population sizes and reduced gene flow, and the incidence of selfing in them is greater (Lawton, 1993; Young *et al.*, 1996; Pujol *et al.*, 2009). The central-marginal hypothesis predicts that peripheral populations have lower genetic diversity and higher differentiation compared to populations in the core of the species' range due to genetic drift, founder effects, inbreeding and bottlenecks (Antonovics *et al.*, 2001; Cole, 2003; Arnaud-Haond *et al.*, 2006; Eckert *et al.*, 2008; Frankham *et al.*, 2010). However, some studies have revealed exceptions to this pattern, including instances of similar (e.g., Putz *et al.*, 2015), and even higher levels of genetic diversity in peripheral populations dependent on population sizes (Lázaro-Nogal *et al.*, 2017). Beyond the general prediction of the central-marginal hypothesis, the genetic diversity and differentiation of peripheral populations can be the result of a complex history, and they may vary greatly depending on the causes behind population origin and on the degree of gene flow with other populations. For example, recent peripheral populations of a species that is currently expanding its range should have low genetic diversity but not show differentiation due to founder effects. Conversely, peripheral populations that have been isolated for a long time (peripheral relict populations) usually exhibit clear genetic differentiation that may be accompanied by a reduced level of genetic variation. In the latter case, the high differentiation makes these populations important for conservation of genetic diversity of the species, and the lower genetic diversity may affect their capacity to adapt to and survive any changes in the evolutionary pressures they experience (Lammi *et al.*, 1999; Cassel-Lundhagen, 2010).

1 In the case of peripheral populations that are isolated from the remaining populations  
2 by long distances, they may be the result of vicariance (fragmentation of an ancient  
3 continuous range, caused by the emergence of barriers to gene flow or by the extinction  
4 of intermediate populations; Rosen, 1978; Wiley, 1988) or long-distance dispersal (LDD)  
5 (Wiley, 1988; Milne, 2006; Cox and Moore, 2016). While LDD is obvious in peripheral  
6 populations occurring on oceanic islands, it may be harder to distinguish vicariance from  
7 LDD in the case of continental populations separated by wide distances without a clear  
8 geographic barrier. The study of phylogenetic relationships and among-population  
9 genetic diversity by using molecular markers can be used to distinguish between these  
10 possibilities (Hewitt, 1999; Avise, 2000; Kropf *et al.*, 2006).

11 The Iberian Peninsula, where European and North African taxa converge, is an area  
12 of high biodiversity (Valdés, 2006; Buirra *et al.*, 2017). Iberian plants related to the  
13 broader European flora have been studied extensively. Their relationship to other  
14 European taxa stems from the fact that the Iberian Peninsula was a refugial area for  
15 temperate plants during the Quaternary ice ages and a source area for subsequent  
16 northward colonisation (Hewitt, 1999). In contrast, studies focused on species distributed  
17 in both Iberia and North Africa and the causes of such distributions are much scarcer  
18 (Caujapé-Castells and Jansen, 2003; Pardo *et al.*, 2008; Terrab *et al.*, 2008; García-Aloy  
19 *et al.*, 2017; Villa-Machío *et al.*, 2018) in spite of the fact that the floras of these two  
20 regions show clear similarities (Valdés, 2006; Fennane and Ibn Tattou, 2012). Dispersal  
21 across the Strait of Gibraltar has been demonstrated to be responsible for the distribution  
22 of some Ibero–North African taxa (Migliore *et al.*, 2012; Lavergne *et al.*, 2013; García-  
23 Aloy *et al.*, 2017) even though this geographical feature has apparently acted as a barrier  
24 to migration or gene flow in many other taxa (e.g. Escudero *et al.*, 2008; Terrab *et al.*,  
25 2008; Casimiro-Soriguer *et al.*, 2010). North Africa is also known to have been the source

1 area for colonisation of the Iberian Peninsula by taxa originating in the Middle East or  
2 distributed in the Mediterranean region (Lumaret *et al.*, 2002; Pardo *et al.*, 2008; Pérez-  
3 Collazos *et al.*, 2009).

4 The genus *Scrophularia* includes approximately 270 species (Ortega-Olivencia and  
5 Devesa, 1993) of primarily Holarctic distribution. The genus originated during the  
6 Miocene (Navarro-Pérez *et al.*, 2013, 2015), probably in Southwest Asia, its primary  
7 centre of diversity (Scheunert and Heubl, 2017). In Europe, the most important centre of  
8 diversity is the Iberian Peninsula, where 12 endemic species and subspecies are present  
9 (Ortega-Olivencia and Devesa, 1993; Ortega-Olivencia, 2009). Iberian endemics are  
10 closely related to those of Morocco (Ortega-Olivencia and Devesa, 2002; Ibn Tattou,  
11 2007) and Macaronesia (Dalgaard, 1979), where five and ten species, respectively, are  
12 also endemic. Phylogenetically, Macaronesian lineages are closely related to western  
13 Mediterranean ones (Scheunert and Heubl, 2014; Navarro-Pérez *et al.*, 2015). One of the  
14 most interesting species is *Scrophularia arguta* Sol. in Aiton, an annual, widespread plant  
15 inhabiting Macaronesia, south-western Europe, North Africa, the Horn of Africa, Arabia  
16 and Socotra Island (Ortega-Olivencia, 2009) (Fig. 1) that is the only known amphicarpic  
17 species in the genus. In Europe, *S. arguta* is restricted to the Iberian Peninsula, where it  
18 is located in two disjunct areas of Spain (Southeast and West-Central).

19 The collection history of *S. arguta* in Europe is unusual, as the number of discovered  
20 populations has increased in the last decades after a long time (nearly 140 years) without  
21 any reference or specimen collected in Europe. The first recorded specimen of this species  
22 in Europe was collected by E. Bourgeau in Sierra de Gádor (Almería Province) on 7 May  
23 1851 and is housed in the herbarium of the Natural History Museum in Paris (Pl. d’Esp.  
24 Exsicc. 1388a, P 89/66P) (see Gay, 1856; Ortega-Olivencia and Devesa, 1993). The  
25 species was considered practically extinct in Spain until Molero (1989) collected it in

1 Águilas (Murcia Province). It was later rediscovered in Sierra de Gádor (Mota *et al.*,  
2 2005). Afterwards, three new populations were found, two in West-Central Spain (Sierra  
3 de Santiago de Alcántara, north-western Cáceres Province; Ortega-Olivencia *et al.*, 2006)  
4 and one in Southeast Spain (southern Granada Province; Cabezudo *et al.*, 2010). Finally,  
5 a few more populations have been discovered in the last few years (see Fig. 1), but always  
6 in Southeast or West-Central Spain. The distance between the westernmost population  
7 (Santiago de Alcántara) and the easternmost population (Mazarrón) is 560 km.

8 All these European populations are peripheral in the context of the widespread  
9 distribution of *S. arguta*. After discovering the West-Central Iberian populations, Ortega-  
10 Olivencia *et al.* (2006) hypothesised that the current disjunct distribution of peripheral  
11 populations of *S. arguta* in the Iberian Peninsula is the result of either vicariance or LDD.  
12 Climate forecasts for the southern half of the Iberian Peninsula predict a future increase  
13 in mean temperature and a decrease in precipitation (Brunet *et al.*, 2009) that could favour  
14 the expansion of Ibero-North African species such as *S. arguta*.

15 Populations in the western part of the range of *S. arguta*, together with a small sample  
16 of eastern populations, were analysed by Valtueña *et al.* (2016) using two nuclear and  
17 two plastid DNA regions. In that study, two Iberian populations of *S. arguta* (one from  
18 the West-Central region and the other from the Southeast) were included. These  
19 peripheral populations had different haplotypes and did not group together in the same  
20 clade, which suggested that they originated from independent colonisation events.  
21 Nevertheless, a more exhaustive study (including more Iberian populations, additional  
22 genetic markers and modelling approaches) is required to determine the origins of  
23 peripheral European populations and estimate their levels of genetic diversity,  
24 differentiation and gene flow. Indeed, the approach of combining genetic data and species

1 distribution modelling is a promising strategy to reconstruct the history of species (e.g.  
2 Fernández-Mazuecos & Vargas, 2013; Leipold *et al.*, 2017; see Morris and Shaw, 2018).

3 Given this background, the main aims of the present study were to: (1) use  
4 phylogeographic analyses (based on nuclear and plastid DNA sequences and AFLP  
5 markers) and species distribution modelling to estimate the temporal and geographical  
6 origin of the peripheral European populations of *S. arguta* and therefore determine the  
7 causes behind their disjunct distribution (vicariance *vs.* LDD), and (2) analyse the genetic  
8 diversity of *S. arguta* populations in the western part of its range to ascertain the  
9 importance of European populations in the context of the global genetic diversity of the  
10 species. With respect to the first objective, we aimed to determine the number, timing and  
11 geographical origin (Northwest Africa and/or Macaronesia) of colonisation events that  
12 gave rise to the Iberian populations. The second objective will provide useful information  
13 to inform conservation policies aimed at maintaining the genetic diversity and  
14 evolutionary potential of this species.

## 15 **2. MATERIALS AND METHODS**

### 16 **2.1. Study species**

17 *Scrophularia arguta* is one of the few annual species and the only amphicarpic species  
18 (i.e., having aerial, chasmogamous flowers as well as basal and/or underground,  
19 cleistogamous ones) in the genus. It is distributed in northern Africa and the Horn, Arabia  
20 and Macaronesia, with a few small, peripheral populations found in south-western Europe  
21 (Iberian Peninsula; Fig. 1), where it is rare. In the Iberian Peninsula, the ecology of south-  
22 eastern populations is different to that of West-Central (Extremadura) populations: the  
23 first set of populations inhabits xeric soils on calcareous or dolomite substrates at low  
24 altitudes (< 400 m asl), whereas the Extremaduran populations inhabit quartzitic rocky

1 areas over 500 m asl. In both cases, plants are found on somewhat nitrified soils at the  
2 foot of shaded walls or between the cracks of these walls.

### 3 **2.2. Sampling strategy**

4 We studied 32 *S. arguta* populations representing most of this species' western  
5 distribution (Fig. 1). The sampling included five populations from the Iberian Peninsula  
6 (i.e., two from Extremadura [West-Central] and one each from the south-eastern  
7 provinces of Granada, Almería and Murcia), 12 from Morocco and 15 from the Canary  
8 Islands (Table 1 and Fig. 1). Three of the Iberian populations were newly sampled for this  
9 study (IB2, IB4 and IB5), while the remaining 29 populations were sampled for previous  
10 studies based on external transcribed spacer (ETS), internal transcribed spacer (ITS) and  
11 ptDNA sequences (Valtueña *et al.*, 2016, 2017). During several collection trips, we  
12 searched for further Iberian populations without success. In addition, four populations  
13 from north-eastern Africa and the Arabian Peninsula were sampled to represent the  
14 eastern part of the species distribution. Twenty-four of the 32 western populations (Table  
15 1) were selected for the generation of amplified fragment length polymorphism (AFLP)  
16 markers, which have not been used to date in this species. Depending on population  
17 abundance, young leaves were collected from one to 20 individuals per population and  
18 stored in silica gel until analysis.

### 19 **2.3. DNA extraction and sequencing**

20 Genomic DNA was extracted using a Qiagen DNeasy Plant Mini kit (Qiagen GmbH,  
21 Hilden, Germany) following the manufacturer's protocol.

22 In a previous study (Valtueña *et al.*, 2016), two nuclear (ITS/ETS) and two plastid  
23 (*psbA-trnH* and *psbJ-petA*) regions were sequenced from 59 individuals and 32  
24 populations of *S. arguta* and one individual of the outgroup species *S. megalantha*  
25 (GenBank accession numbers: KU945636–KU945694 for ETS, KC692563 and



1 KU926629–KU926686 for ITS, KU945695–KU945754 for *psbA-trnH* and KU945755–  
2 KU945815 for *psbJ-petA*). For this study, the same four regions were sequenced in two  
3 individuals of each newly sampled population (i.e., 24 new sequences). Amplification of  
4 these regions was performed as described by Valtueña *et al.* (2016). Sequencing was  
5 carried out in both strand directions by the Service of Applied Techniques for Biosciences  
6 (SATB) of Extremadura University (Badajoz, Spain). Sequences were manually checked  
7 and edited using Sequencher v4.10 (GeneCodes, Ann Arbor, MI, USA) and aligned by  
8 eye with MacClade v4.08 (Maddison and Maddison, 2005). All new sequences obtained  
9 in this study were submitted to GenBank (see Table S1).

#### 10 **2.4. Sequences analyses**

11 In the previous investigation (Valtueña *et al.*, 2016), sequences were analysed using  
12 statistical parsimony in TCS (Clement *et al.*, 2000) to obtain a haplotype network, and  
13 using Bayesian inference in BEAST 1.8.2 (Drummond *et al.*, 2012) to determine  
14 phylogenetic relationships of populations. These analyses were repeated using the same  
15 settings and calculation procedures but including the three newly sampled Iberian  
16 populations.

17 The absence of resolution at the base of the *S. arguta* tree and the radial pattern of the  
18 haplotype network obtained in the previous study (Valtueña *et al.*, 2016) were interpreted  
19 as the result of demographic expansion, and the date of differentiation was estimated to  
20 be c. 3.28 Ma ago. In the present study, we therefore performed a mismatch distribution  
21 analysis (MDA) of combined ptDNA sequences of all sampled *S. arguta* populations to  
22 confirm the demographic expansion. To further verify the expansion model, we compared  
23 the raggedness index (HRag; Harpending, 1994) and goodness of fit based on the sum of  
24 squared deviations (SSD) between observed and expected mismatch distributions. Small  
25 values of HRag support sudden population expansion whereas high values suggest

1 stationary or bottlenecked populations (Harpending *et al.*, 1993; Harpending, 1994). The  
2 date of a confirmed expansion (t) can be estimated according to the formula  $t = \tau/2\mu k$ ,  
3 where  $\tau$  is the expansion parameter,  $\mu$  is the mutation rate per site and generation, and k  
4 is the sequence length. The analysis was performed in Arlequin 3.5.2.2 (Excoffier and  
5 Lischer, 2010) with the same ptDNA matrix used in the TCS analysis after removing the  
6 outgroup taxon *S. megalantha* (to avoid overestimation of the number of mutations and  
7 the value of  $\tau$ , complex indels were treated as single mutations). The assumed mutation  
8 rate based on plastid noncoding regions was  $1.52 \times 10^{-9}$  substitutions per site per year  
9 (Yamane *et al.*, 2006), and the sequence length was 1142 bp.

10 We reconstructed historical migration routes of *S. arguta* using Bayesian discrete  
11 phylogeographic analysis (DPA) (Lemey *et al.*, 2009). ITS/ETS and ptDNA sequences  
12 were analysed separately. Analyses were conducted in BEAST v1.8.4. (Drummond *et al.*,  
13 2012). We defined four areas (Canary Islands, Iberian Peninsula, Northwest Africa and  
14 Northeast Africa) and assigned each *S. arguta* sample to one of them. *S. megalantha* was  
15 included as outgroup (with undetermined distribution), and all ingroup (*S. arguta*)  
16 sequences were constrained as a monophyletic group. Substitution models were selected  
17 using jModeltest (Darriba *et al.*, 2012), and areas were mapped under an asymmetric  
18 substitution model. To simplify the analyses and facilitate convergence, a strict molecular  
19 clock (with rates as specified earlier) and a constant-size coalescent tree prior were  
20 implemented. For each analysis, two MCMC analyses were run for 10 million  
21 generations, sampling every 1000 generations. Analysis with Tracer (Rabaut and  
22 Drummond, 2007) confirmed adequate sample sizes. The two chains were combined  
23 using LogCombiner (Drummond *et al.*, 2012) after discarding the first 10% of sampled  
24 generations as burn-in, and trees were summarized in a MCC tree using TreeAnnotator  
25 (Drummond *et al.*, 2012).

## 1 2.5. AFLPs

2 For the AFLP analysis, we used 305 individuals belonging to 24 populations (populations  
3 with only one to three sampled individuals were excluded; Table 1). All reactions were  
4 performed simultaneously for all samples in three steps. Restriction-ligation and  
5 preselective amplification steps followed the protocol of Kropf *et al.* (2002), except that  
6 30 cycles were used instead of 20 cycles in the preselective PCR program. The selective  
7 amplification followed that of Trybusch *et al.* (2006). Adapters and primers used in each  
8 step are listed in Table S2. AFLP products were separated as a multiplex using primers  
9 labelled with different fluorescent dyes (6-Fam, Hex and 56-TAMN) by the SATB.  
10 Reproducibility of the results was confirmed using replicate extracts of 14 samples.

11 AFLP fragments were scored manually with GeneMarker 1.5 (SoftGenetics, State  
12 College, PA, USA) for the presence/absence of fragments between 70 and 450 bp in size.  
13 Fragments that could not be scored unambiguously were excluded. The results of the  
14 scoring were converted to a presence/absence (i.e., 1/0) matrix and used for further  
15 analysis. Because all mismatched fragments among sample replicates were removed from  
16 the matrix, the matrix error rate was zero.

17 The genetic structure of the dataset was analysed using BAPS v6.0 (Corander *et al.*,  
18 2008). The analysis was run 20 times for  $K = 2$  to 24, where  $K$  is the maximum number  
19 of populations assumed to be present in the sample. Admixture analyses (Corander and  
20 Marttinen, 2006) were run with 100 iterations to estimate admixture coefficients for  
21 individuals, with 200 reference individuals from each population and 20 iterations used  
22 to estimate admixture coefficients for reference individuals. In addition to this analysis, a  
23 spatial genetic mixture analysis of groups (Cheng *et al.*, 2013) was performed in which  
24 the populations and population coordinates were incorporated into the analysis and  $K$  was  
25 set to 24. We additionally implemented Bayesian clustering in STRUCTURE (Pritchard

1 *et al.*, 2000; Falush *et al.*, 2003) with a number of clusters  $K = 1$  to 24 (number of sampled  
2 populations). For each  $K$ , ten independent chains were run with 1,000,000 MCMC  
3 replicates after 100,000 burn-in replicates, admixture model and independent allele  
4 frequencies among populations. Post-processing was carried out with CLUMPAK  
5 (Kopelman *et al.*, 2015) to compare  $K$  values, using Delta  $K$  (Evanno *et al.*, 2005) and  
6 Prob( $K$ ), and to summarise results. In addition, to confirm genetic groups found in the  
7 BAPS and STRUCTURE analyses, a principal coordinates analysis (PCoA), a non-  
8 hierarchical grouping technique not dependent on prior knowledge of the source location  
9 of the sampled individuals, was conducted in GenAlEx v6.502 (Peakall and Smouse,  
10 2006).

11 Various diversity parameters (number of fragments, number of private fragments,  
12 percentage of polymorphic loci and heterozygosity [ $H_E$ ]) were calculated in GenAlEx at  
13 both population and group levels (groups being phylogenetic clades based on the  
14 combined analysis in BEAST –which are the same clades found in the DPA analysis of  
15 the plastid regions– and main haplogroups found in the TCS analysis). Gene diversity  
16 (total gene diversity [ $H_T$ ], within-population gene diversity [ $H_S$ ] and genetic  
17 differentiation [ $F_{ST}$ ]) at the group level (phylogenetic clades and main haplogroups) were  
18 calculated in PopGene 32 (Yeh *et al.*, 2000). The distribution of genetic variation in *S.*  
19 *arguta* was assessed by analysis of molecular variance (AMOVA) as implemented in  
20 Arlequin 3.5.2.2 (Excoffier and Lischer, 2010). The AMOVA was run with 1000  
21 permutations considering the whole dataset (24 populations), with two and three  
22 hierarchical levels (i.e., considering the five phylogenetic clades and the seven main  
23 haplogroups).

## 24 **2.6. Species distribution modelling**

1 We used species distribution modelling (SDM) to evaluate the potential range of *S. arguta*  
2 in the present and to project it to late Quaternary conditions, including the last interglacial  
3 (LIG, c. 120–140 ka), the last glacial maximum (LGM, c. 22 ka) and the mid-Holocene  
4 (MH, c. 6 ka). The analysis was focused on our study region, including the Iberian  
5 Peninsula, Northwest Africa and the Canary Islands (26° to 45° N; 20° W to 5° E). Highly  
6 accurate geographic coordinates of 35 localities of *S. arguta* in this region were obtained  
7 from our sampled populations and herbarium specimens. The scarcity of occurrence data  
8 in eastern Africa prevented us from including this area. To reduce sampling bias, the  
9 dataset was filtered by randomly removing points that were within a buffer of 0.2°,  
10 resulting in a final dataset of 23 occurrences.

11 Layers of 19 bioclimatic variables under present conditions at a resolution of 30'' were  
12 obtained from WorldClim 1.4. To avoid collinearity, we excluded variables displaying a  
13 high correlation with other variables ( $|r| > 0.7$ ) in the study region. As a result, a set of  
14 seven variables were selected: bio1, bio3, bio4, bio6, bio12, bio14 and bio15. SDM was  
15 performed using the maximum entropy algorithm, implemented in Maxent v3.4 (Phillips  
16 *et al.*, 2006). We used 80% of occurrences for model training, and 20% for model  
17 evaluation. Ten subsample replicates were run, and a mean model was calculated. The  
18 model was projected to past conditions using LIG layers from Otto-Bliesner *et al.* (2008)  
19 and LGM and MH layers based on the CMIP5 project (three global climate models  
20 [GCMs]: CCSM4, MIROC-ESM and MPI-ESM-P). Resolution was 30'' for LIG and MH  
21 layers, and 2.5' for LGM layers. For LGM and MH, an average of the three GCMs was  
22 calculated.

23

## 24 **3. RESULTS**

### 25 **3.1. Sequences analyses**

1 Iberian populations did not cluster together in the combined nuclear and plastid Bayesian  
2 tree (Fig. S1). Five main clades were identified in the tree (posterior probability = 0.99–  
3 1.00), but their relationships were unsupported. Two of these clades included Canarian  
4 populations only (Fig. S1): one comprising populations from the three westernmost  
5 islands sampled (western Canarian clade, WCa), and the other including populations from  
6 the two easternmost islands (eastern Canarian clade, ECa). Iberian populations were  
7 included in the remaining three clades. The two West-Central Iberian populations (IB1  
8 and IB2) constituted their own clade (Extremaduran clade, Ex). The two south-eastern  
9 populations (IB3 and IB4, from Murcia and Almeria provinces, respectively) were  
10 included in a clade together with populations from Morocco (located on the Atlantic coast  
11 and the central Moroccan interior) and eastern populations (Arabia, Sudan and Socotra  
12 Island; widespread clade, Wid clade). Finally, the south-eastern Iberian population closest  
13 to Morocco (IB5, Granada Province) grouped with northern Moroccan populations  
14 located along the Mediterranean coast (North Morocco-Iberian clade, Mo-Gr).

15 Two of the three newly analysed Iberian populations had the same haplotypes as their  
16 nearest previously studied Iberian neighbour. In particular, population IB2 shared  
17 haplotype C with population IB1, and population IB4 shared haplotype A6 with  
18 population IB3 (Table 1 and Fig. 1). In contrast, population IB5 showed a haplotype that  
19 was not found in our previous study. This haplotype (F2) was only one mutational step  
20 away from haplotype F1 (haplotype F in our previous study). Haplotype F1 was only  
21 found in population MO3 (Table 1 and Fig. 1), located near the Moroccan coast across  
22 from population IB5.

23 The results of the MDA supported a sudden expansion model ( $HR_{ag} = 0.01062$ ,  $p =$   
24  $0.518$ ;  $SSD = 0.00768$ ,  $p = 0.417$ ). The value of  $\tau$  was 12.34378, with the expansion  
25 accordingly dated to 3.55 Ma.

1 DPAs for ptDNA and ITS/ETS sequences recovered some uncertainty on the location  
2 of the most recent common ancestor of all *S. arguta* samples, with similar posterior  
3 probabilities for the Canary Islands and Northwest Africa, and lower values for the  
4 remaining two areas (Figs. 2 and S2). In the ptDNA analysis (Fig. 2), five major clades  
5 were recovered, matching those of previous analyses. Two of these clades had a most  
6 probable ancestral location in the Canary Islands, two in Northwest Africa and one in the  
7 Iberian Peninsula. Within one of the ancestrally Northwest African clades (Wid), the  
8 analysis inferred migrations to Northeast Africa (two events), the Canary Islands (one  
9 event) and the Iberian Peninsula (one event, populations IB3 and IB4). A second probable  
10 migration to the Iberian Peninsula from Northwest Africa was inferred in the Mo-Gr clade  
11 (population IB5). A third migration to the Iberian Peninsula was suggested (populations  
12 IB1 and IB2), but its origin remained inconclusive. The analysis estimated that all  
13 migration events to the Iberian Peninsula occurred during the Quaternary, specifically in  
14 the last 2 million years. The ITS/ETS analysis was hardly informative due to the low  
15 probabilities of most ancestors (Fig. S2), but it was consistent with multiple origins of  
16 Iberian populations, including a supported Northwest African origin of population IB5.

### 17 **3.2. AFLPs**

18 The six AFLP primers generated 492 fragments. The mean number of fragments per  
19 individual was  $122.0 \pm 16.2$  (range: 66–157).

20 The best partition uncovered in the BAPS analysis was at  $K = 13$ . However, at  $K = 9$ ,  
21 the rate of change in the log probability of the data between successive  $K$  values was  
22 stabilised, which is considered to be a better indicator of the number of clusters (Fig. S3).  
23 In both cases ( $K = 13$ , Fig. 3A; and  $K = 9$ , Fig. S4), Iberian populations were included in  
24 three different clusters. Populations IB1 and IB2 formed a clearly differentiated cluster  
25 that appeared at  $K = 3$  (Fig. S4). Populations IB3 and IB4 formed an exclusive cluster at

1  $K = 13$  (Fig. 3A), but they were clustered with MO8 at lower values of  $K$  (Fig. S4).  
2 Finally, population IB5 clustered with population MO3 and a few individuals of  
3 population MO4 at  $K = 13$  (Fig. 3A), with population MO2 also included in this group at  
4  $K = 9$  (Fig. S4). In all Iberian populations, all individuals from a given population were  
5 assigned to the same cluster, as confirmed by the admixture analysis (Fig 3B). The best  
6 partition obtained from spatial clustering of groups of individuals was at  $K = 11$   
7 ( $\log[\text{marginal likelihood}] = -42963.2343$ ). Populations IB1 and IB2 formed a cluster, as  
8 did populations IB3 and IB4 (Fig. 3C). Population IB5 shared a cluster with populations  
9 MO2 and MO3 (Fig. 3C). In STRUCTURE analyses, Prob( $K$ ) suggested  $K = 19$  as the  
10 optimal clustering ( $\text{Prob}[K] = 1$ ), while Delta  $K$  displayed the highest value at  $K = 3$ , with  
11 a further small peak at  $K = 9$  (Fig. S3C). Results for  $K = 9$ , which were considered the  
12 most informative for our purposes, are shown in Fig. 3D whereas values from  $K = 2$  to  $K$   
13  $= 12$  are shown in Fig S5. The clustering was similar to that recovered from BAPS, but  
14 with a higher degree of admixture. Iberian populations were again recovered in three  
15 different clusters. Populations from West-Central Iberian Peninsula (IB1 and IB2) formed  
16 a cluster on their own, while populations from Southeast Iberia were clustered with North  
17 African populations. Specifically, populations IB3 and IB4 were clustered with a  
18 population from southern Morocco (MO8) while population IB5 was clustered with  
19 Moroccan populations located just across the Mediterranean Sea (MO2, MO3 and MO4)  
20 (Fig. 3D).

21 In the PCoA analysis, the first two axes explained 27.2% of the variance, with this  
22 percentage increasing to 33.4% when the third axis was considered. These results  
23 confirmed the clear differentiation of populations IB1 and IB2 with respect to the  
24 remaining populations (Fig. 4). When only the first two axes were considered, individuals  
25 from populations IB3 and IB4 clustered together with individuals from population MO8,



1 whereas individuals from population IB5 overlapped with those of populations MO2,  
2 MO3 and MO4 (Fig. 4A).

3 Values of genetic parameters calculated from the AFLP data at population,  
4 phylogenetic clade and main haplogroup levels are shown in Table 2. At the population  
5 level, the number of fragments varied between 173 (MO3) and 304 (MO10). The  
6 percentage of polymorphic loci ranged between 19.51% (LA7) and 59.55% (MO10),  
7 while  $H_E$  varied between 0.059 (LA7) and 0.152 (MO10). Only four populations showed  
8 a private fragment (IB1, TE1, TE2 and MO2). Iberian populations had intermediate  
9 values of the three parameters, with population IB1 displaying the highest number of  
10 fragments and IB4 the highest percentage of polymorphic loci and heterozygosity. When  
11 the phylogenetic clades uncovered by the Bayesian analysis were considered, the lowest  
12 number of fragments (276) was found in the Ex and WCa clades, with the former also  
13 exhibiting the lowest number of polymorphic loci (46.34%) and heterozygosity (0.120).

14 Gene diversity and differentiation values at the level of species, phylogenetic clades  
15 and main haplogroups are shown in Table 3. The  $H_S$  index (reflecting within-population  
16 gene diversity) at the species level was 0.108 and the  $H_T$  index (total gene diversity) was  
17 0.222, while genetic differentiation among populations was very high ( $F_{ST} = 0.515$ ).  
18 When phylogenetic clades were considered,  $H_S$  varied between 0.089 (ECa clade) and  
19 0.124 (Wid clade), and  $H_T$  ranged from 0.119 (Ex clade) to 0.185 (Wid clade). The highest  
20 genetic differentiation was found in the ECa clade ( $F_{ST} = 0.400$ ) and the lowest in the Ex  
21 clade ( $F_{ST} = 0.075$ ). With respect to haplogroups,  $H_S$  varied between 0.089 (haplogroup  
22 D) and 0.128 (E), and  $H_T$  ranged from 0.119 (C) to 0.193 (E). The highest genetic  
23 differentiation was found in haplogroup D ( $F_{ST} = 0.400$ ) and the lowest in haplogroup C  
24 ( $F_{ST} = 0.075$ ).

1 According to the results of the AMOVA when populations were not grouped into  
2 clusters, genetic variation was similar among and within populations (53.6% vs. 46.4%,  
3 respectively; Table 4). When phylogenetic clades and main haplogroups were considered,  
4 the genetic variation found within populations was similar (43.7% vs. 44.5%). The genetic  
5 variation among groups and among populations was almost the same for haplogroups  
6 (27.9% vs. 27.6%); in the case of phylogenetic clades, however, the genetic variation  
7 found among groups was clearly higher than among populations (30.4% vs. 25.9%).

### 8 **3.3. Species distribution modelling**

9 Areas climatically suitable for *S. arguta* in the present were inferred mostly in western  
10 Morocco, north-eastern Morocco, north-western Algeria and the Canary Islands (Fig. 5).  
11 Narrow suitable areas were also inferred in the southern coast of the Iberian Peninsula,  
12 including known localities in Southeast Spain. However, localities in West-Central Spain  
13 (Extremadura) displayed low suitability. Projections to the past (Fig. 5) revealed stable  
14 suitable areas throughout the LIG, LGM and MH in western Morocco, the Canary Islands  
15 and, to a lesser degree, north-eastern Morocco. Low suitability in the Iberian Peninsula  
16 was inferred for the LIG, but suitable areas appear to have been present in south-eastern  
17 Spain at least since the LGM. Low suitability was estimated in West-Central Spain for all  
18 periods.

## 19 **4. DISCUSSION**

### 20 **4.1. Genetic structure of western *Scrophularia arguta* populations**

21 In a previous study, the last common ancestor of *Scrophularia arguta* lineages was dated  
22 to the late Pliocene (c. 3.28 Ma; Valtueña *et al.*, 2016) based on two secondary calibration  
23 points. In the current study, mismatch distribution analysis confirmed this result but gave  
24 a slightly older date (3.55 Ma). The DNA sequences (Figs. 1, 2, S1) and AFLP data (Figs.  
25 3, 4; Tables 2-4) revealed several genetic lineages in this species that mainly

1 corresponded to different geographical regions. The same result was obtained by  
2 Valtueña *et al.* (2016), who used ITS/ETS and plastid DNA sequences only, and was  
3 explained by the life-history traits of *S. arguta*: annual habit and reproductive success  
4 largely dependent on the production of cleistogamous flowers. The westernmost portion  
5 of the range of this species encompasses Macaronesia, Northwest Africa and the Iberian  
6 Peninsula, where several genetic lineages with distinct origins are found (see below).

7 With respect to Macaronesia, Valtueña *et al.* (2016) using plastid markers and  
8 focusing on the Canary Islands uncovered two ancient lineages with distinct distributions  
9 on the islands. Those findings were confirmed in the present study by the AFLP clustering  
10 analysis. The presence of ancient endemic lineages is also consistent with the long-term  
11 persistence of suitable areas on the Canary Islands in the Quaternary, as indicated by  
12 SDM (Fig. 5). The present genetic analyses additionally suggested a close relationship  
13 between some populations of Lanzarote (ECa clade) and Atlantic Moroccan coastal  
14 populations at low values of  $K$ . This suggests the existence of long-distance gene flow  
15 between the two areas (Atlantic Moroccan coast and Lanzarote). Long-distance gene flow  
16 has been previously proposed for other species exhibiting clear differentiation based on  
17 plastid DNA (Besnard *et al.*, 2007; Migliore *et al.*, 2012). Gene flow between both areas  
18 is supported by the STRUCTURE results, where most of the individuals from population  
19 MO1 show an important probability to belong to a genetic group constituted by Lanzarote  
20 populations at  $K = 9$  (Fig. 3D). This result indeed implies a directionality of gene flow  
21 from the island to mainland. Gene flow may have been favoured during the LGM by  
22 marine regression that reduced the distance between both areas (Fernández-Palacios *et*  
23 *al.*, 2011).

24 Phylogenetic analysis indicated the existence of two clades in Northwest Africa, one  
25 of which included populations from the eastern range of the species (Wid clade; Figs. 2,

1 S1). The exact biogeographic relationship between eastern and western populations is still  
2 unclear. While the haplotype network suggests that haplotype A colonised Morocco from  
3 the east and was the founder of the Mo-Gr clade (see Valtueña *et al.*, 2016), DPA analysis  
4 estimates that eastern populations most probably originated from western populations  
5 (Fig. 2). In any case, the high genetic diversity of *S. arguta* in Morocco seems to be due  
6 to *in situ* differentiation probably as result of climatic fluctuations over the last three  
7 million years (Lavergne *et al.*, 2013) that favoured differentiation in isolated areas rather  
8 than multiple waves of colonisation from the eastern range of the species or recolonisation  
9 from Europe/Macaronesia. This is consistent with the long-term climatic suitability in  
10 Northwest Africa estimated by SDM (Fig. 5) and supports the idea that this area acted as  
11 a refuge for numerous taxa during the Pleistocene (Rodríguez-Sánchez *et al.*, 2008;  
12 Désamoré *et al.*, 2011; García-Aloy *et al.*, 2017). The major lineages of *S. arguta* most  
13 likely arose by differentiation in isolated refugial areas during unfavourable periods when  
14 populations experienced range reductions. This contrasts with the situation in  
15 Macaronesia and Europe, where the multiple genetic groups of *S. arguta* are explained  
16 by several independent colonisation events.

#### 17 **4.2. Three independent origins of peripheral European populations**

18 Both the nucleotide and AFLP data (Figs. 1-4) support at least three jumps onto the  
19 Iberian Peninsula involving three different geographic areas: (a) the province of Granada,  
20 (b) Southeast Iberia (Murcia and Almería provinces) and (c) West-Central Iberia  
21 (Extremadura). In addition, the results of the AFLP analysis demonstrate the absence of  
22 gene flow among the three areas. Gene flow was not detected even between the two  
23 geographically closest areas (196 km between IB4 and IB5).

24 Following the discovery of the Extremaduran populations, the disjunct distribution of  
25 *S. arguta* in the Iberian Peninsula was suggested to be a consequence of either vicariance

1 or LDD (Ortega-Olivencia *et al.*, 2006). Valtueña *et al.* (2016) supported the hypothesis  
2 of two different colonisation events (two populations analysed, one from each Iberian  
3 area, fell into different clades). Our latest results, including genetic analyses and SDM,  
4 reject the vicariance hypothesis, and indicate that three colonisation events, rather than  
5 two, have taken place. Although repeated waves of colonization have been frequently  
6 demonstrated in other geographical contexts (e.g. Reisch, 2008; Escudero *et al.*, 2010),  
7 only two herbaceous species are currently known to have colonised the Iberian Peninsula  
8 from Northwest Africa more than once: the annual *Hypochaeris glabra* and the perennial  
9 *H. radicata* (Ortíz *et al.*, 2008, 2009). In both of these cases, two colonisation events were  
10 inferred. Our study is the first to demonstrate the occurrence of at least three different  
11 colonisation events. Although *S. arguta* does not apparently have a syndrome for LDD,  
12 this species exhibits some life-history traits (annual habit, self-compatible breeding  
13 system, cleistogamous flowers, small seeds [dust-like in chasmogamous flowers] and a  
14 preference for somewhat disturbed habitats) that are compatible with this mode of  
15 dispersal (Lavergne *et al.*, 2013; García-Aloy *et al.*, 2017).

16 The Granadian population (IB5) is closely related to coastal Mediterranean  
17 populations of Morocco, as these populations formed a clade in the phylogenetic analysis  
18 and appeared in the same genetic group in the AFLP clustering analyses. Both results and  
19 potential areas estimated by SDM support the establishment of the IB5 population as a  
20 result of a relatively recent dispersal. The absence of AFLP differentiation from  
21 Mediterranean Moroccan populations suggests a late Quaternary dispersal from this area,  
22 probably the most recent dispersal event involving the Iberian Peninsula (see also DPA  
23 results in Fig. 2). Being located at the edge of the sea, containing few individuals and  
24 being restricted to a few meters of limestone-dolomitic rocky coast, the Granadian  
25 population is in a precarious situation: it may become extinct if the area suffers any

1 disturbances due to local anthropic activity or to the sea level rise induced by global  
2 climate change that is obviously occurring in the Mediterranean Sea as well as worldwide  
3 (Galassi and Spada, 2014; Zerbini *et al.*, 2017).

4 Interestingly, our analyses revealed that the Murcian–Almerian populations (IB3,  
5 IB4) are closely related to those from West-Central Morocco. Two hypotheses might  
6 explain this relationship: (1) LDD from West-Central Morocco or (2) dispersal from  
7 northern Morocco (LDD or stepping stone dispersal), with the ancestral population of IB3  
8 and IB4 subsequently becoming extinct, overrun by individuals with other genotypes, or  
9 not sampled by us. This second hypothesis is more likely, and the extinction or  
10 displacement of the ancestral population is supported by the fact that the Northwest  
11 African populations have suffered waves of contraction and expansion in response to  
12 climatic fluctuations in the Quaternary, as shown by SDM results (Fig. 5; Médail and  
13 Diadema, 2009; Hussemann *et al.*, 2014). Under either scenario, the colonisation of the  
14 south-eastern peninsula by individuals with haplogroup A must have predated the  
15 colonisation that gave rise to the Granadian population (IB5), as IB3 and IB4 populations  
16 are genetically different from their most closely related Moroccan populations. This  
17 colonisation event could have used the large island remnants of a volcanic arc in the East  
18 Alboran basin existing until the Early Pleistocene (located to the East of the Strait of  
19 Gibraltar; Booth-Rea *et al.*, 2018) as stepping stones between northern Morocco and  
20 south-eastern Spain.

21 According to the AFLP data (Table 2), the IB3 population has a lower number of  
22 fragments, percentage of polymorphic loci and heterozygosity, possibly a consequence  
23 of a founder effect if that population has recently originated from IB4. This low genetic  
24 variation may alternatively indicate that IB3 is suffering a bottleneck. This is supported  
25 by the low number of individuals present, i.e., an average of 33 mature individuals per

1 year (BORM, 2015) with annual fluctuations between 4 and 85. In 2012, when samples  
2 were collected, fewer than 20 individuals were present.

### 3 **4.3. A peripheral relict population in West-Central Iberia**

4 The Extremaduran populations (IB1, IB2) represent a well-differentiated lineage within  
5 *S. arguta* based on four lines of evidence. First, they are not grouped with any other  
6 populations in the phylogenetic tree (Figs. 2, S1). Second, they possess a unique plastid  
7 haplotype that differs by at least seven mutational steps from the closest haplotypes in  
8 any other population (Fig. 1). Third, they constitute an independent genetic group based  
9 on the AFLP clustering analyses, in which they formed the first group to be differentiated  
10 (beginning at  $K = 3$ ; Figs. 3, 4, S4). And fourth, some degree of ecological differentiation  
11 exists in these populations, as their distribution is not predicted by SDM for any of the  
12 analysed periods (Fig. 5). In addition, the two populations have similar genetic  
13 characteristics; they share most identified AFLP loci and show low differentiation ( $F_{ST} =$   
14  $0.075$ ; Table 3). Considering these results together with the close proximity between the  
15 two populations (0.628 km), these seem to behave as a single population or be the result  
16 of recent fragmentation of a larger population. The high genetic differentiation between  
17 these plants and the remaining range of *S. arguta* for all molecular markers supports that  
18 the dispersal event that originated these populations is the oldest involving the Iberian  
19 Peninsula, predating the other two colonisations of Iberia uncovered in the present study  
20 (see Fig. 2).

21 Our results provide support for the idea that the Extremaduran populations constitute  
22 an evolutionary significant unit. In addition, given the location of these populations at the  
23 northern limit of the species range, they can be considered peripheral relict populations  
24 (Cassel-Lundhagen, 2010). In the haplotype network (Fig. 1), the haplotype of  
25 Extremaduran populations and those of the eastern Canarian populations share a common

1 ancestor, which is directly connected to the ancestral haplotype of the Wid clade (A1).  
2 However, given the low support for phylogenetic relationships of the Ex clade, the DPA  
3 was unable to confidently infer the origin of Extremaduran populations (Fig. 2), with the  
4 Canary Islands and Northwest Africa being the most likely options. The latter hypothesis  
5 seems more likely, as it implies a smaller dispersal distance and is consistent with the  
6 waves of contraction and expansion of Northwest African populations that may have led  
7 to the disappearance of ancestral genotypes, similar to the scenario described for the  
8 ancestor of Murcian–Almerian populations. The present-day range of *S. arguta* in West-  
9 Central Iberia is very restricted, and its habitat is completely different to that of other *S.*  
10 *arguta* populations, as supported by the low suitability in this area estimated by the SDM  
11 analysis for all periods. Their genetic parameters, however, do not show any evidence of  
12 negative effects caused by a reduced distribution and the lack of gene flow with other  
13 populations. This may be due to the fact that the two populations collectively have a large  
14 number of individuals (c. 4591 in 2006, at least 1285 in 2017 and c. 1000 in 2018), an  
15 atypical situation for this species.

#### 16 **4.4. The importance of peripheral populations for conservation**

17 Interestingly, the history of European populations of *S. arguta* is very diverse, and it is  
18 not possible to refer to them as a single set. Each group of populations found in our study  
19 corresponds to a different type of peripheral population. The Granadian population (IB5)  
20 would be a peripheral population of an expanding species, and it has recently been  
21 originated by dispersal. As a result, genetic characteristics of this population are modelled  
22 by founder effect (low genetic diversity and no genetic differentiation from source  
23 populations; Schwaegerle and Schaal, 1979; Arnaud-Haond, 2006). Murcian–Almerian  
24 populations (IB3 and IB4) have been isolated from their source populations for a longer  
25 period of time, which has allowed the accumulation of mutations leading to some genetic



1 differentiation. In this case, a founder effect is not detected, and low genetic diversity is  
2 probably due to bottlenecks. Finally, Extremaduran populations (IB1 and IB2) are  
3 peripheral relict populations, displaying a strong differentiation from the rest of the  
4 populations of the species and a similar genetic diversity (Cassel-Lundhagen, 2010; Plenk  
5 *et al.*, 2017). Overall, the European populations of *S. arguta* are a clear example of the  
6 evolution of peripheral populations in absence of gene flow, with population IB5  
7 corresponding to the first stages after dispersal, populations IB1 and IB2 to populations  
8 isolated for a prolonged period of time, and populations IB3 and IB4 to an intermediate  
9 situation.

10 Therefore, the conservation value of the Iberian populations of *S. arguta* is  
11 unquestionable, given the idiosyncrasies of the three population groups. For example,  
12 populations in the south of the Iberian Peninsula probably contain alleles related to  
13 tolerance to water stress, while the Extremaduran populations may contain alleles related  
14 to adaptation to more mesic environments. Extremaduran plants are particularly relevant  
15 for conservation, since they make up one of the five major intraspecific clades on their  
16 own, and their extinction would mean the disappearance of genetic variants that are found  
17 nowhere else in the distribution range of *S. arguta*. The singularity of these populations  
18 reflects the importance of ancient peripheral populations for the global genetic diversity  
19 of species and highlights the need to understand the history of peripheral populations in  
20 order to conserve intraspecific diversity.

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**Table 1.** Studied populations of *Scrophularia arguta*. Population abbreviations (Pop), locations, haplotypes (HAP), phylogenetic clades (PHY), numbers of individuals used (N) in the AFLP analysis and BAPS groups considering  $K = 13$  (Group) are indicated –: no individuals used in the AFLP analysis

Pop	Location	HAP	PHY	AFLP	
				N	Group
Iberian Peninsula (Spain):					
IB1	Spain, Cáceres, Santiago de Alcántara	C	Ex	20	1
IB2	Spain, Cáceres, Santiago de Alcántara	C	Ex	14	1
IB3	Spain, Murcia, Águilas	A <sub>6</sub>	Wid	8	2
IB4	Spain, Almería, Pulpí	A <sub>6</sub>	Wid	14	2
IB5	Spain, Granada, Almuñécar	F <sub>2</sub>	Mo-Gr	9	3
Canary Islands (Spain):					
FU1	Fuerteventura, Tetir	D <sub>1</sub>	ECa	13	4
FU2	Fuerteventura, Tiscamanita	D <sub>1</sub>	ECa		–
FU3	Fuerteventura, Joros	D <sub>1</sub>	ECa		–
GC	Gran Canaria, La Isleta	A <sub>2</sub>	Wid		–
GO	La Gomera, Barranco de Guarimiar	G <sub>2</sub>	WCa		–
LA1	Lanzarote, Tahiche	D <sub>2</sub>	ECa	18	5
LA2	Lanzarote, Punta de las Mujeres	D <sub>2</sub>	ECa	14	5
LA3	Lanzarote, Jameos del Agua	D <sub>3</sub>	ECa	10	4/5
LA4	Lanzarote, Orzola	D <sub>1</sub>	ECa		–
LA5	Lanzarote, San Bartolomé	D <sub>1</sub>	ECa	12	4
LA6	Lanzarote, Tinajo	D <sub>1</sub>	ECa	18	4
LA7	Lanzarote, El Golfo	D <sub>3</sub>	ECa	15	4
PA	La Palma, Santa Cruz	G <sub>2</sub>	WCa		–
TE1	Tenerife, Güimar	G <sub>1</sub>	WCa	13	6
TE2	Tenerife, Pal-Mar	G <sub>2</sub>	WCa	13	6/7
Northwestern Africa:					
MO1	Morocco, Safi Cape	E <sub>1</sub>	Wid	13	8/9
MO2	Morocco, Zegangane	E <sub>3</sub>	Mo-Gr	13	10
MO3	Morocco, Hassi-Berkane	F <sub>1</sub>	Mo-Gr	5	3
MO4	Morocco, Had-Rouadi	E <sub>3</sub>	Mo-Gr	15	3/11
MO5	Morocco, Beni-Sidel	E <sub>3</sub>	Mo-Gr	15	9/11/12
MO6	Morocco, Sidi-Bou-Othmane	A <sub>5</sub>	Wid	4	12
MO7	Morocco, Oued El-Abid Gorges	E <sub>2</sub> /E	Wid	10	11/12/13
		1			
MO8	Morocco, Ouzaghar	A <sub>2</sub>	Wid	12	13
MO9	Morocco, Oued Assaka	A <sub>2</sub>	Wid	12	11/12/13
MO10	Morocco, Beddouza	B	Wid	15	9/11/12
MO11	Morocco, Safi	E <sub>1</sub>	Wid		–
MO12	Morocco, Jebel Agouti, Agadir Melloul	A <sub>2</sub>	Wid		–
Northeastern Africa and Arabian Peninsula:					
SA1	Saudi Arabia, Jabal Hada	A <sub>4</sub>	Wid		–
SA2	Saudi Arabia, Al-Baha	A <sub>4</sub>	Wid		–
SO	Yemen: Socotra, Fiheri Park	A <sub>1</sub>	Wid		–
SU	Sudan: Arkawit, Jebel Elsit	A <sub>3</sub>	Wid		–

**Table 2.** Genetic parameters (number of alleles, NA; number of private alleles, NPA; percentage of polymorphic loci, PPL; and expected heterozygosity,  $H_E$ ) of *Scrophularia arguta* at population, phylogenetic clade and main haplogroup levels. The number of populations and studied individuals constituting each phylogenetic clade or main haplogroup are indicated in parentheses. Population abbreviations are as in Table 1

	NA	NPA	PPL (%)	$H_E$
<b>Populations</b>				
IB1	242	1	36.59	0.107
IB2	238	0	36.38	0.112
IB3	187	0	25.20	0.084
IB4	232	0	42.28	0.122
IB5	201	0	32.93	0.097
FU1	189	0	21.75	0.062
LA1	230	0	32.93	0.101
LA2	239	0	35.47	0.102
LA3	234	0	36.99	0.114
LA5	200	0	32.11	0.102
LA6	210	0	29.47	0.081
LA7	184	0	19.51	0.059
TE1	190	1	29.07	0.090
TE2	233	1	42.48	0.123
MO1	273	0	49.80	0.140
MO2	180	1	23.58	0.073
MO3	173	0	22.56	0.082
MO4	256	0	47.56	0.143
MO5	285	0	54.07	0.143
MO6	178	0	24.39	0.090
MO7	269	0	51.02	0.141
MO8	233	0	42.07	0.125
MO9	277	0	55.08	0.134
MO10	304	0	59.55	0.152
Mean $\pm$ SD	226.5 $\pm$ 37.7	0.17 $\pm$ 0.38	36.79 $\pm$ 11.58	0.107 $\pm$ 0.027
<b>Phylogenetic clades</b>				
Ex clade (2/34)	276	7	46.34	0.120
Wid clade (8/88)	452	5	91.87	0.177
Mo-Gr clade (5/57)	382	8	76.42	0.165
ECa clade (7/100)	348	4	67.48	0.149
WCa clade (2/26)	276	2	53.25	0.143
<b>Haplogroups</b>				
A (5/50)	388	1	78.86	0.168
B (1/15)	304	0	59.55	0.152
C (2/34)	276	7	46.34	0.120
D (7/100)	348	4	67.48	0.149
E (5/66)	433	1	87.20	0.179
F (2/14)	239	0	44.11	0.125
G (2/26)	276	2	53.25	0.143

**Table 3.** AFLP total ( $H_T$ ) and within-population ( $H_S$ ) gene diversity, and genetic differentiation ( $F_{ST}$ ) for the entire dataset, phylogenetic clades and main haplogroups with more than one population

	$H_S$	$H_T$	$F_{ST}$
Species (all populations)	0.108	0.222	0.515
Phylogenetic clades			
Ex clade	0.110	0.119	0.075
Wid clade	0.124	0.185	0.330
Mo-Gr clade	0.108	0.170	0.368
ECa clade	0.089	0.148	0.400
WCa clade	0.110	0.145	0.244
Haplogroups			
A	0.111	0.175	0.365
C	0.110	0.119	0.075
D	0.089	0.148	0.400
E	0.128	0.193	0.336
F	0.090	0.127	0.291
G	0.110	0.145	0.244

1



**Table 4.** Analysis of molecular variance of *Scrophularia arguta* AFLP data considering the entire dataset (24 populations) with two and three hierarchical levels (both for five phylogenetic clades and seven main haplogroups)

Source of variation	d.f.	Sum of squares	Variance components	% of variation	<i>P</i>
Populations not grouping					
Among populations	23	10976.435	477.236	53.6	0.001
Within populations	281	8596.909	30.594	46.4	0.001
Phylogenetic clades					
Among groups	4	6063.799	1515.950	30.4	0.000
Among populations	19	4869.234	256.275	25.9	0.000
Within populations	281	8640.312	30.748	43.7	0.000
Main haplogroups					
Among groups	6	6361.008	1060.168	27.9	0.000
Among populations	17	4572.025	268.943	27.6	0.000
Within populations	281	8640.312	30.748	44.5	0.000

1

1 Figure 1. Distribution map of studied populations of *Scrophularia arguta* from the  
2 western range of the species and TCS statistical parsimony network of plastid DNA  
3 haplotypes. Colours on the map and the haplotype network correspond to the main  
4 haplogroups. Phylogenetic clades indicated on the haplotype network are represented on  
5 the map by the following symbols: circle, Widespread clade (Wid); diamond,  
6 Extremaduran clade (Ex); hexagon, western Canarian clade (WCa); square, North  
7 Moroccan–Granadian clade (Mo–Gr); triangle, eastern Canarian clade (ECa). White stars  
8 indicate Iberian populations not sampled in this study. Population abbreviations are the  
9 same as in Table 1. In the haplotype network, the dashed line indicates the connection to  
10 the outgroup (*S. megalantha*), and haplotypes included in Iberian populations are  
11 encircled by a solid line. Small circles represent inferred mutational steps (white, one  
12 step; grey, more than one step, as indicated). Haplotype abbreviations are as in Table 1.  
13 Inset: map showing global distribution of the species.

14 Figure 2. Discrete phylogeographic analysis (DPA) of *Scrophularia arguta* based on  
15 ptDNA sequences. The maximum clade credibility tree is shown, with branches coloured  
16 according to the most probable ancestral range. Branch thickness indicates posterior  
17 probability (PP), with PPs>0.90 also shown as numbers above branches. Pie charts  
18 represent PP distributions of ancestral range at well supported (PP≥0.95) nodes.

19 Figure 3. BAPS and STRUCTURE clustering analyses of a *Scrophularia arguta* AFLP  
20 dataset. (A) BAPS result for  $K = 13$ , (B) BAPS admixture result for  $K = 13$ , (C) BAPS  
21 result from the spatial clustering analysis, and (D) STRUCTURE result for  $K = 9$ .  
22 Population abbreviations are as in Table 1.

23 Figure 4. Principal coordinates analysis of the AFLP dataset of *Scrophularia arguta*  
24 based on genetic distances. Percentages of total variance explained by the first three  
25 coordinates are shown on the respective axes. Sample abbreviations are as in Table 1.

1 Ellipses indicate supported clades in the phylogenetic analysis (clade abbreviations as in  
2 Fig. 1). Inset in B shows the eigen values for the first 10 axes.

3 **Figure 5.** Species distribution modelling (SDM) of *Scrophularia arguta*, including the  
4 average model fitted to present climatic conditions and average projections to the last  
5 interglacial (c. 120–140 ka), last glacial maximum (c. 22 ka) and mid-Holocene (c. 6 ka).  
6 The logistic output of the Maxent model is shown. Localities used to build the model are  
7 shown as red dots in the present map.